

Synthesis and Structure-activity Relationships of a Novel Oral Carbapenem, CS-834

MASAO MIYAUCHI, ROKURO ENDO, MASAFUMI HISAOKA, HIROSHI YASUDA
and ISAO KAWAMOTO*

Research Laboratories, Sankyo Co., Ltd.,
1-2-58 Hiromachi, Shinagawaku, Tokyo 140, Japan

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We have studied an ester prodrug of a carbapenem to develop a potent orally active β -lactam antibiotic. A variety of 1β -methylcarbapenem derivatives have been synthesized. We have found that some derivatives having an amide group in the C-2 side chain show potent and well balanced antibacterial activities as well as high stability against dehydropeptidase-I. Oral absorption of derivatives has been optimized by modifying the C-3 ester promoiety. Pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate, CS-834, has been selected as the most promising compound for further evaluation.

An orally active antibiotic with potent activity is of much interest in the clinical realm because oral administration and lower dosage are advantageous for patients.

Carbapenems are the most potent β -lactam antibiotics which have a broad antibacterial spectrum and potent bactericidal activities against both Gram-positive and -negative organisms.^{1,2)} They are highly resistant to hydrolysis by a variety of β -lactamases. So far, imipenem,³⁾ panipenem⁴⁾ and meropenem⁵⁾ have been launched on the market, and several compounds are currently under clinical evaluation.^{6~8)} However, most compounds have been developed for parenteral use, and none for the practical purpose of oral administration. Currently, tricyclic β -lactam antibiotics, GV-104326 and its ester GV-118819, have been developed. GV-118819 is now under clinical study as an oral anti-infective drug.^{9~11)}

The chemical instability of carbapenem has been recognized as a serious problem for the development of oral carbapenem, although 1β -methyl substitution of carbapenem improved its stability to some extent.¹²⁾ We expected that 1β -methylcarbapenems could be stable enough for oral administration by preferable contribution of the 1β -methyl group, and that the dosage could be reduced as a result of their powerful antibacterial effects.

We have synthesized a variety of 1β -methylcarbapenems derivatized at the C-2 side chain and evaluated them for antibacterial activities and other biological properties. We have found that some derivatives having an amide group in the C-2 side chain show potent and

well balanced antibacterial activity as well as high stability against dehydropeptidase-I. These compounds have been submitted for prodrug approach to optimize oral absorption. Through the investigation we have successfully obtained an orally active carbapenem, pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (CS-834).¹³⁾

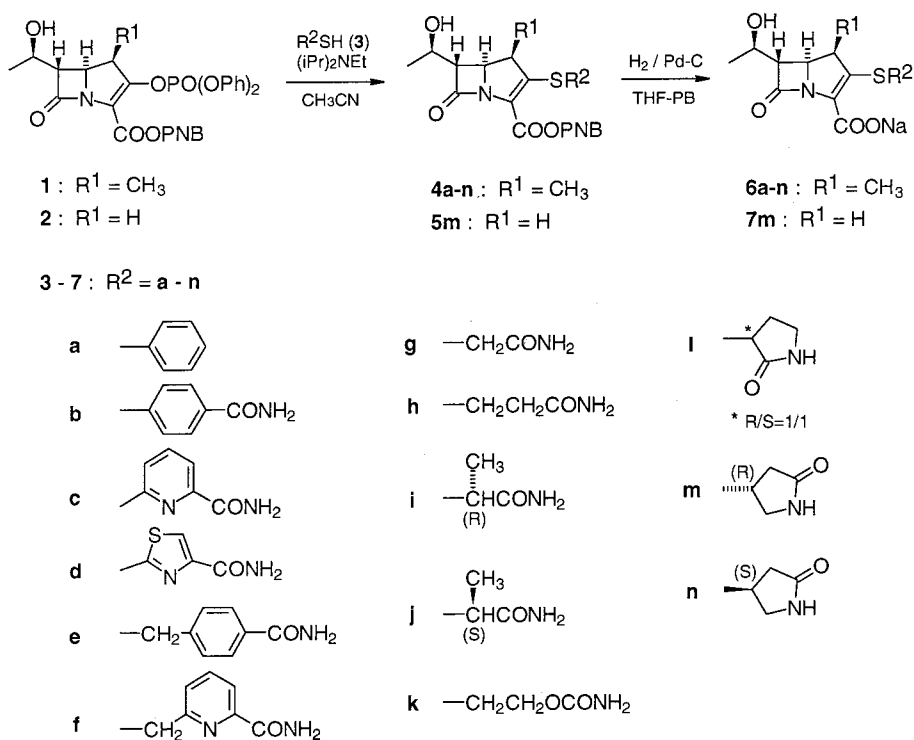
In this paper we will describe the synthesis and structure-activity relationship of CS-834 and related compounds.

Chemistry

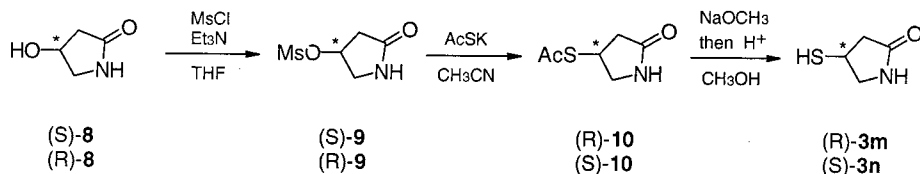
The C-2 substituted derivatives of carbapenems were synthesized through the general procedure shown in Scheme 1. It includes condensation of the carbapenem-2-yl diphenylphosphate **1**^{12,14)} with appropriate thiol **3a~n** designed for the C-2 side chain to give carbapenem ester **4a~n**, and the following deprotection of the 4-nitrobenzyl group by catalytic hydrogenation using 10% palladium on charcoal. The resulting carbapenem derivatives **6a~n** were purified by reversed phase column chromatography and lyophilized as sodium salts. The 1-H carbapenem **7m**, which has the same C-2 side chain as the derivative **6m**, was also prepared in a similar way.^{15,16)} Synthesis of derivatives **6a~6l** will be reported elsewhere. The sodium salt **6m** (R-83201) was converted to the corresponding free acid (R-95867) by treatment with 1 N HCl in water.

Optically active (*R*)- and (*S*)-4-mercapto-2-oxopyrrolidine **3m** and **3n** were prepared starting from (*S*)- and (*R*)-4-hydroxy-2-oxopyrrolidine **8**,¹⁷⁾ respectively, as

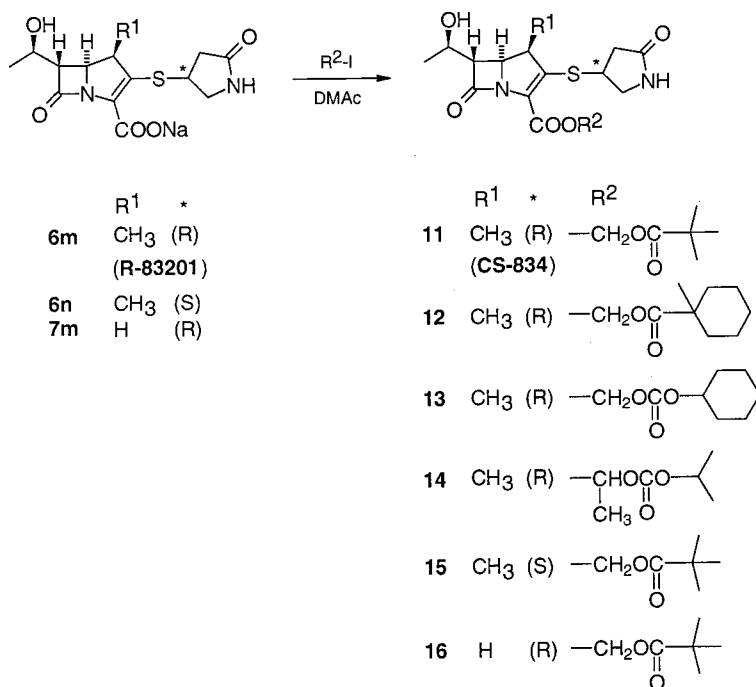
Scheme 1.



Scheme 2.



Scheme 3.



shown in Scheme 2.

Several acyloxyalkyl and alkoxy-carbonyloxyalkyl groups were selected as potential ester promoieties with effective absorption from the intestinal tract following hydrolysis to the parent compound. Ester prodrugs **11**~**16** were prepared from carbapenems **6m**, **6n** and **7m** in order to give them enough lipophilicity for oral absorption as shown in Scheme 3.

Among the ester derivatives **11**~**16**, pivaloyloxy-methyl ester of the (*R*)-isomer **11** was obtained as a crystalline form, although the pivaloyloxymethyl ester of the (*S*)-isomer **15** was an amorphous powder. The crystalline form of **11** is of greater value for practical use because it is stable in storage and easy to handle and formulate.

The chemical stability of the ester in solution is also an important factor because an orally administered compound dissolves in the gastrointestinal fluid prior to absorption. The chemical stability of carbapenem ester **11** was examined in a phosphate buffer solution of pH 6.86 at 37°C. The carbapenem **11** decomposed apparently according to first order kinetics at a rate constant of 0.031 hour⁻¹.¹⁸⁾ The carbapenem showed higher stability than various cephalosporins, of which we have examined the chemical stability under similar conditions.¹⁹⁾ This finding is contrary to the conventional view that carbapenems are unstable even at neutral pH.

Biological Properties

The antimicrobial activity (MICs) of carbapenems are shown in Tables 1 and 2.

First of all, the antimicrobial activity of aromatic and heteroaromatic derivatives which have a carboxamide group in the C-2 side chain were examined. The carbapenem **6a**, which has only a phenylthio group, showed excellent activity against Gram-positive bacteria such as *Staphylococcus aureus*, but it was inactive against Gram-negative bacteria. However, the aromatic and heteroaromatic derivatives **6b**~**f** having a carboxamide group showed improved activity against Gram-negative bacteria compared to **6a** as well as excellent activity against Gram-positive bacteria. The aryl and heteroarylmethylthio derivatives **6e** and **6f** were more active against Gram-negative bacteria than the corresponding aryl and heteroarylthiol compounds **6b** and **6c**, respectively.

Secondly, aliphatic carboxamide or carbamate derivatives were examined. The derivatives **6g**~**j** having an alkyl carboxamide were potent against Gram-negative bacteria, but they were less active against Gram-positive bacteria. The stereochemistry of the methyl group on the carbon next to the sulfur atom influenced the antibacterial activity. The (*R*)-isomer **6i** showed better antibacterial activity than the (*S*)-isomer **6j**. The carbamate derivative **6k** showed potent activity similar to the carboxamides **6g** and **6h**.

Finally, in order to promote higher activity against Gram-positive bacteria, a 5-membered cyclic amide system was introduced. Although 2-oxopyrrolidin-3-ylthio derivative **6l** became less active, the 5-oxopyrrolidin-3-ylthio derivatives **6m** and **6n** exhibited well balanced and potent antimicrobial activity against

Table 1. Antibacterial activity (MIC, µg/ml)^a of carbapenems **6a**~**k** and urinary recovery (%)^b in mice.

	6a	6b	6c	6d	6e	6f	6g	6h	6i	6j	6k
<i>Staphylococcus aureus</i> 209P	≤0.01	≤0.01	≤0.01	≤0.01	0.02	0.02	0.1	0.1	0.1	0.2	0.05
<i>S. aureus</i> 56R	0.05	≤0.01	0.02	0.02	0.05	0.05	0.2	0.1	0.1	0.2	0.1
<i>S. aureus</i> 535 (MRSA)	3.1	1.5	3.1	1.5	3.1	3.1	12.5	12.5	50	50	25
<i>Enterococcus faecalis</i> 681	6.2	6.2	6.2	3.1	0.8	0.8	3.1	3.1	1.5	12.5	1.5
<i>Escherichia coli</i> NIHJ	25	1.5	3.1	0.8	0.4	0.8	0.05	0.02	≤0.01	0.1	0.05
<i>E. coli</i> 609	25	6.2	6.2	3.1	1.5	1.5	0.1	0.05	0.05	0.1	0.1
<i>Salmonella enteritidis</i>	6.2	0.8	1.5	0.4	0.2	0.2	0.05	0.02	≤0.01	0.1	0.02
<i>Klebsiella pneumoniae</i> 806	25	3.1	3.1	0.8	0.4	0.8	0.05	0.02	≤0.01	0.1	0.02
<i>K. pneumoniae</i> 846	12.5	1.5	12.5	1.5	3.1	12.5	0.05	0.05	0.02	0.4	0.4
<i>Enterobacter cloacae</i> 903	25	6.2	50	>12.5	12.5	12.5	0.8	1.5	0.8	1.5	3.1
<i>Serratia marcescens</i> 1184	25	3.1	6.2	1.5	0.8	3.1	0.1	0.05	0.02	0.2	0.1
<i>Proteus vulgaris</i> 1420	0.2	0.05	3.1	0.1	0.1	0.1	0.4	0.1	0.02	0.2	0.1
<i>Morganella morganii</i> 1510	6.2	1.5	1.5	1.5	0.4	0.4	0.4	0.4	0.4	0.4	0.2
<i>Pseudomonas aeruginosa</i> 1001	100	>12.5	25	>12.5	>100	200	25	50	50	50	50
Urinary recovery (%) (s.c.)	33	N.T.	26	N.T.	9	41	47	40	45	72	32

^a MIC was determined by agar dilution method with an inoculum of 10⁷ cfu/ml.

^b Urinary recovery (%) was determined by disk method using *Bacillus subtilis* ATCC 6633 as a test strain after subcutaneous administration of compound (50 mg/kg) in SPF ddY mice (n=5, 0~24 hours).

Table 2. Antibacterial activity (MIC, $\mu\text{g/ml}$)^a of carbapenems **6l**~**n** and **7m** and urinary recovery (%)^b in mice.

	6l	6m (R-83201)	6n	7m	GV 104326 ^c	CPDX
<i>Staphylococcus aureus</i> 209P	0.2	0.02	0.05	≤ 0.01	0.02	0.8
<i>S. aureus</i> 56R	0.4	0.02	0.1	≤ 0.01	0.05	0.8
<i>S. aureus</i> 535 (MRSA)	25	6.2	12.5	6.2	12.5	>200
<i>Enterococcus faecalis</i> 681	12.5	1.5	3.1	0.8	0.8	25
<i>Escherichia coli</i> NIHJ	0.1	≤ 0.01	≤ 0.01	≤ 0.01	0.2	0.4
<i>E. coli</i> 609	0.2	0.02	0.05	0.02	1.5	0.4
<i>Salmonella enteritidis</i>	0.1	≤ 0.01	0.02	0.02	0.2	0.1
<i>Klebsiella pneumoniae</i> 806	0.1	≤ 0.01	≤ 0.01	≤ 0.01	0.4	0.1
<i>K. pneumoniae</i> 846	1.5	0.02	0.05	0.05	1.5	0.8
<i>Enterobacter cloacae</i> 903	6.2	0.8	0.8	1.5	6.2	3.1
<i>Serratia marcescens</i> 1184	0.4	≤ 0.01	0.05	0.02	0.8	0.2
<i>Proteus vulgaris</i> 1420	0.2	0.02	0.2	0.2	0.4	≤ 0.01
<i>Morganella morganii</i> 1510	0.4	0.1	0.2	0.2	0.8	100
<i>Pseudomonas aeruginosa</i> 1001	50	25	50	50	100	>200
Urinary recovery (%) (s.c.)	55	75	46	8	87	91

^a MIC was determined by agar dilution method with an inoculum of 10^7 cfu/ml.

^b Urinary recovery (%) was determined by disk method using *Bacillus subtilis* ATCC 6633 as a test strain after subcutaneous administration of compound (50 mg/kg) in SPF ddY mice (n=5, 0~24 hours).

^c GV104326 was synthesised according to Glaxo's patent procedure.

both Gram-positive and -negative bacteria. As for the configuration at the C-3 position of 5-oxopyrrolidine, the (*R*)-isomer **6m** was slightly more active than the (*S*)-isomer **6n** against *S. aureus*, *E. faecalis* and some Gram-negative bacteria.

The 1-H carbapenem **7m** which has the same C-2 side chain as compound **6m** showed similar activity to **6m**.

The antimicrobial activity of **6m** is superior to that of cefpodoxime (CPDX)²⁰⁾ and almost equal to that of parenteral carbapenems such as imipenem and panipenem. However, they are less active against *Pseudomonas aeruginosa*. A new tricyclic β -lactam, GV-104326, seems to be inferior to compound **6m** in antibacterial activity against Gram-negative bacteria.

The urinary recovery of carbapenems was determined after subcutaneous administration to mice in order to estimate stability of carbapenems under biological conditions. Derivatives having an alkyl or cyclic amide were recovered in moderate amounts (40~75%) as shown in Tables 1 and 2.

Some stereoselective effects of the chiral center in the C-2 side chain were observed in urinary recovery. As to the alkyl amides **6i** and **6j**, recovery of the (*R*)-isomer **6j** was less than that of the (*S*)-isomer **6i** although **6j** had better antibacterial activity. With respect to the cyclic amides **6m** and **6n**, urinary recovery was highly dependent on the configuration at the C-3 position of 5-oxopyrrolidine. The recovery of the (*R*)-isomer **6m** was 75%, although the recovery of the (*S*)-isomer **6n** was only 46%.

Table 3. Effect of cilastatin on urinary recovery of carbapenems in mice.

Compound	Urinary recovery ^a (%)		DHP-I Degradation rate ratio ^b
	CS (-)	CS (+)	
1 β -Methylcarbapenem			
6m (<i>R</i>)(R-83201)	75	84	1.0
6n (<i>S</i>)	46	78	3.3
1-H Carbapenem			
7m (<i>R</i>)	8	64	14.2

^a Carbapenems (50 mg/kg) were subcutaneously administered to mice (n=5) with or without cilastatin (CS: 50 mg/kg as sodium salt). Urinary recovery was determined by the same method described in Table 1.

^b Mouse renal dehydropeptidase-I (DHP-I) was used. The figures are relative to the degradation rate of **6m**.

Simultaneous administration of cilastatin, an inhibitor of dehydropeptidase-I, with the (*S*)-isomer **6n** resulted in improved urinary recovery equal to that of the (*R*)-isomer **6m**. In addition, the degradation rate of **6m** with mouse dehydropeptidase-I (DHP-I) was slower than that of **6n** as listed in Table 3. These results clearly demonstrate that the (*R*)-isomer **6m** is reliable in the biological system due to higher resistance against DHP-I in mice.²¹⁾

The recovery of the 1-H carbapenem **7m** was only 8%, which was in accordance with the fast degradation by DHP-I as listed in Table 3. The urinary recovery of **7m** was increased to 64% by coadministration with cilastatin. This result shows that the 1-H carbapenem **7m**

is very labile in mice due to degradation by DHP-I.

From these experiments the 1 β -methylcarbapenem **6m** which has an (*R*)-5-oxopyrrolidin-3-ylthio substitution at the C-2 position has been selected as the best parent compound for the following prodrug approach.

In order to optimize the oral absorption of the carbapenem **6m**, several ester derivatives were prepared as described above. The oral absorption of a prodrug can be estimated from its urinary recovery as the parent compound. The urinary recoveries after oral administration of prodrugs to mice are shown in Table 4. Improved urinary recovery was observed in all of the derivatives (**11** ~ **14**). Among them, especially, the pivaloyloxymethyl

ester **11** and the 1-methylcyclohexylcarbonyloxymethyl ester **12** had excellent urinary recoveries of 47% and 55%, respectively. Pivaloyloxymethyl ester **15** prepared from the (*S*)-isomer **6n** showed lower urinary recovery than ester **11** of the (*R*)-isomer **6m**. These results reflect the better biological stability of the (*R*)-isomer.

The pivaloyloxymethyl ester **16** of the 1-H carbapenem **7m** was hardly recovered in urine probably due to the chemical and biological instability of **7m** and **16**.

In order to find the best ester promoity, the pharmacokinetics of prodrugs **11** and **12** were compared with each other in dogs. Results are listed in Table 5. The pivaloyloxymethyl ester **11** showed better profiles than the 1-methylcyclohexylcarbonyloxymethyl ester **12**. The maximum plasma level (C_{max}) for **11** was 6.1 $\mu\text{g/ml}$ and the plasma half life ($T_{1/2}$) was 0.90 hours at an oral dose of 10 mg/kg as the parent carbapenem **6m**. The absolute oral bioavailability of **11** was approximately twice that of **12**.

Based on good oral absorption and potent antibacterial activity, the prodrug **11** showed excellent therapeutic efficacy against systemic infections in mice. It was highly effective especially against infections caused by resistant strains of bacteria, to which oral cephalosporins had been inactive.²²⁾

Through these experiments the pivaloyloxymethyl ester **11** was selected as the most promising compound for further evaluation.

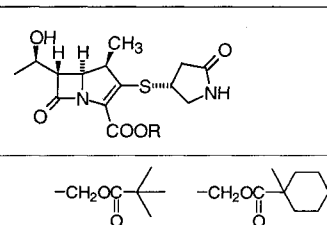
Table 4. Urinary recovery of ester derivatives after oral administration in mice.

Compound	Urinary recovery ^a (%)
1 β -Methylcarbapenem	
11 (<i>R</i>)(CS-834)	46
12 (<i>R</i>)	55
13 (<i>R</i>)	44
14 (<i>R</i>)	21
15 (<i>S</i>)	24
1-H Carbapenem	
16 (<i>R</i>)	0.3
GV-118819 ^b	41
CPDX-PR	67

^a Compounds (50 mg/kg) were orally administered to mice (n = 5) and urinary recovery was determined by the same method described in Table 1.

^b GV-118819 was synthesized according to Glaxo's patent procedure.

Table 5. Pharmacokinetic parameters of carbapenems in dogs (Beagle, n = 3) after intravenous injection of **6m** or oral administration of its esters **11** and **12**^a.

R			
	6m (R-83201)	11 (CS-834)	12
C_{max} ($\mu\text{g/ml}$)	38.4 (at 5 min)	6.1	2.6
T_{max} (hour)	—	1.00	0.83
$T_{1/2}$ (hour)	0.75	0.90	0.94
AUC ($\mu\text{g}\cdot\text{hr/ml}$)	26.3	13.1	6.3
Absolute bioavailability (%)	100	50	24

^a The parent compound (**6m**) was intravenously injected to dogs (Beagle, n = 3) at a dose of 10 mg/kg. Esters (**11** and **12**) were orally administered at a dose of 10 mg/kg as **6m**.

Conclusion

We have successfully obtained an oral carbapenem, pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(1*R*)-1-hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate **11** (CS-834) which shows potent and well balanced antibacterial activity and both chemical and biological reliability. CS-834 is now under clinical evaluation.

Experimental

General Methods

IR spectra were recorded on a Nicolet NIC FT-IR (5SXC) spectrometer. NMR spectra were determined on a Jeol GX-270 (270 MHz) or GX-400 (400 MHz) spectrometer using tetramethylsilane (TMS) or sodium 3-(trimethylsilyl)propionate-*d*₄ (TSP) as an internal standard. The mp was determined using a Yanagimoto micro-melting point apparatus and was not corrected. UV spectra were recorded on a Shimadzu UV-3100 spectrometer. Optical rotations were determined on a Perkin-Elmer digital polarimeter. Column chromatography was carried out on Silica gel 60 (230~400 mesh, Art.9385, Merck), Cosmosil 75C₁₈ PREP (75 μm, Nacalai tesque) or MCI GEL CHP-20P (75~150 μm, Mistubishi Kasei Corporation).

Preparation of (*R*)-4-Mercapto-2-oxopyrrolidine (**3m**)

i) (*S*)-4-Methanesulfonyloxy-2-oxopyrrolidine **9**

To a solution of (*S*)-4-hydroxy-2-oxopyrrolidine **8** (1.90 g, 18.8 mmol) in pyridine (100 ml), methanesulfonyl chloride (2.26 g, 19.7 mmol) was added dropwise under ice-cooling. The mixture was stirred at room temperature for 1.5 hours, after which the reaction mixture was concentrated by evaporation under reduced pressure. Aqueous NaHCO₃ was then added to the mixture, and the mixture was again concentrated to dryness by evaporation under reduced pressure. A mixture of EtOAc-MeOH (1:1) was then added to the resulting residue. The insoluble portion was removed by filtration and the soluble portion was concentrated by evaporation under reduced pressure. The residue obtained from the soluble portion was chromatographed through silica gel (Merck 9385, 150 ml) with EtOAc-MeOH (from 9:1 to 4:1) as the eluent to give a crystalline residue. The residue was recrystallized from EtOAc-MeOH to obtain the compound **9** (2.44 g, 13.6 mmol, 72% yield) as crystals; mp 137.5~139°C; $[\alpha]_D^{24} -35.5^\circ$ (*c* 1.09, MeOH); IR (KBr) cm⁻¹ 1719, 1697, 1659, 1305, 1177,

1171, 1159, 963; ¹H NMR (400 MHz, DMSO-*d*₆, TMS) δ 2.28 (1H, dd, *J*=17.6 and 1.8 Hz), 2.71 (1H, dd, *J*=17.6 and 6.3 Hz), 3.24 (3H, s), 3.37 (1H, d, *J*=11.9 Hz), 3.66 (1H, dd, *J*=11.9 and 5.3 Hz), 5.31~5.34 (1H, m), 7.85 (1H, br s).

ii) (*R*)-4-Acetylthio-2-oxopyrrolidine **10**

To a solution of the compound (*S*)-**9** (896 mg, 5.00 mmol) in CH₃CN (90 ml) was added potassium thioacetate (857 mg, 7.50 mmol), and the mixture was then refluxed for 2 hours. Insoluble material was removed by filtration, and the filtrate was concentrated by evaporation under reduced pressure. The residue was chromatographed on a silica gel column with EtOAc-MeOH (from 1:0 to 96:4) as the eluent to give a crystalline residue (593 mg). The residue was recrystallized from EtOAc-cyclohexane to give the compound (*R*)-**10** as crystals (455 mg, 2.85 mmol, 57% yield); mp 59~60°C; $[\alpha]_D^{25} +47.3^\circ$ (*c* 1.33, MeOH); IR (KBr) cm⁻¹ 1689, 1125; ¹H NMR (400 MHz, CDCl₃, TMS) δ 2.30 (1H, dd, *J*=17.4 and 6.0 Hz), 2.35 (3H, s), 2.80 (1H, dd, *J*=17.4 and 9.1 Hz), 3.31 (1H, dd, *J*=10.2 and 5.1 Hz), 3.89 (1H, dd, *J*=10.2 and 7.2 Hz), 4.15~4.23 (1H, m), 7.27 (1H, br s).

iii) (*R*)-4-Mercapto-2-oxopyrrolidine **3m**

To a solution of the compound (*R*)-**10** (375 mg, 2.35 mmol) in MeOH (5 ml) was added 1*N* NaOMe in MeOH (2.35 ml, 2.35 mmol) under ice-cooling, and the mixture was stirred for 20 minutes. To the mixture was added 1*N* aq. HCl (2.35 ml, 2.35 mmol), and then the mixture was concentrated to dryness by evaporation under reduced pressure. EtOAc was added to the residue and insoluble matter was removed by filtration. The filtrate was concentrated by evaporation under reduced pressure to give the title compound **3m** as crystals (275 mg, 2.34 mmol, 100% yield); mp 69.5~70°C; $[\alpha]_D^{25} +36.5^\circ$ (*c* 1.18, MeOH); IR (KBr) cm⁻¹ 2539, 1699, 1683; ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.96 (1H, d, *J*=7.2 Hz), 2.32 (1H, dd, *J*=17.2 and 6.8 Hz), 2.80 (1H, dd, *J*=17.2 and 8.2 Hz), 3.32 (1H, dd, *J*=10.2 and 5.6 Hz), 3.62~3.70 (1H, m), 3.81 (1H, dd, *J*=10.2 and 7.3 Hz), 7.27 (1H, br s).

(*S*)-4-Mercapto-2-oxopyrrolidine (**3n**)

The (*S*)-isomer **3n** was prepared from (*R*)-4-hydroxy-2-oxopyrrolidine **8** as described for the preparation of the (*R*)-isomer **3m**. MP, IR and ¹H NMR spectra of the (*S*)-isomer **3n** were identical to those of the (*R*)-isomer **3m**. $[\alpha]_D^{25} -36.8^\circ$ (*c* 1.04, MeOH).

Preparation of Sodium (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**6m**, R-83201)

i) 4-Nitrobenzyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate **4m**

To a solution of 4-nitrobenzyl (1*R*,5*S*,6*S*)-2-diphenylphosphoryloxy-6-[(*R*)-1-hydroxyethyl]-1-methyl-1-carbapen-2-em-3-carboxylate **1** (1000 mg, 1.68 mmol) was added a solution of the mercaptan **3m** (197 mg, 1.68 mmol) in CH₃CN and diisopropylethylamine (296 μ l, 1.68 mmol). The mixture was stirred for 1 hour under ice-cooling and then left to stand overnight at 4°C. The crystalline compound which precipitated from the reaction mixture was collected by filtration and dried to give the compound **4m** as crystals (672 mg, 1.45 mmol, 86% yield); mp 219~221°C;

Anal Calcd for C₂₁H₂₃N₃O₇S: C 54.66, H 5.02, N 9.11.

Found: C 54.72, H 5.12, N 8.97.

IR (KBr) cm⁻¹ 1756, 1684, 1521, 1338, 1148; ¹H NMR (270 MHz, DMSO-*d*₆, TMS) δ 1.16 (3H, d, *J*=6.3 Hz), 1.17 (3H, d, *J*=7.3 Hz), 2.13 (1H, dd, *J*=17.1 and 4.4 Hz), 2.79 (1H, dd, *J*=17.1 and 7.8 Hz), 3.10 (1H, dd, *J*=10.8 and 3.4 Hz), 3.16~3.35 (1H, m), 3.40~3.51 (1H, m), 3.70 (1H, dd, *J*=10.7 and 7.3 Hz), 3.95~4.12 (2H, m), 4.25 (1H, dd, *J*=9.3 and 2.5 Hz), 5.07 (1H, d, *J*=5.4 Hz), 5.30 (1H, d, *J*=14.2 Hz), 5.46 (1H, d, *J*=14.2 Hz), 7.71 (2H, d, *J*=8.8 Hz), 8.23 (2H, d, *J*=8.8 Hz).

ii) Sodium (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate **6m**

To a solution of the compound **4m** (390 mg, 0.78 mmol) in a mixture of THF (19 ml) and 0.1 M aq. phosphate buffer (pH 7.0) (18 ml) was added 10% Pd-charcoal (300 mg), and the mixture was stirred vigorously at 30°C in an atmosphere of hydrogen for 2.5 hours. The catalyst was removed by filtration from the reaction mixture, and the filtrate was washed twice with Et₂O. The aqueous layer was then concentrated by evaporation under reduced pressure, and the resulting residue was chromatographed through MCI GEL CHP-20P (Mitsubishi Kasei Corporation, 75~150 μ m, 50 ml) developed with water. The desired fraction was concentrated to 10 ml, and then lyophilized to give the title compound **6m** as a colorless powder (225 mg, 0.65 mmol, 83% yield);

Anal Calcd for C₁₄H₁₇N₂O₅SNa·2H₂O:

C 43.75, H 5.51, N 7.29.

Found: C 43.31, H 5.25, N 7.25.

IR (KBr) cm⁻¹ 1749, 1687, 1596, 1393, 1295; UV (H₂O) nm 299; ¹H NMR (270 MHz, D₂O, TSP) δ 1.02 (3H, d, *J*=7.3 Hz), 1.10 (3H, d, *J*=6.6 Hz), 2.22 (1H, dd, *J*=17.6 and 4.4 Hz), 2.77 (1H, dd, *J*=17.6 and 8.4 Hz), 3.08~3.25 (2H, m), 3.25 (1H, dd, *J*=5.9 and 2.6 Hz), 3.68 (1H, dd, *J*=11.4 and 6.4 Hz), 3.84~3.96 (1H, m), 4.00~4.12 (2H, m).

Preparation of (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylic acid (R-95867)

To a solution of the sodium salt **6m** (500 mg, 1.44 mmol) in water (2.5 ml) was added 1 N HCl (1.44 ml, 1.44 mmol) under ice-cooling, and then the mixture was stirred for 30 minutes. The crystal which precipitated from the mixture was taken by filtration and dried to give the title compound (446 mg, 1.37 mmol, 95% yield); mp 204~206°C;

Anal Calcd for C₁₄H₁₈N₂O₅S·H₂O:

C 48.83, H 5.85, N 8.13, S 9.31.

Found: C 48.78, H 5.83, N 7.99, S 9.29.

IR (KBr) cm⁻¹ 3502, 3464, 3293, 3143, 3033, 2980, 1782, 1698, 1650, 1545, 1487, 1450, 1416, 1377, 1281, 1258, 1226, 1201, 1183, 1141, 1128, 1101, 1083, 1069, 1047; ¹H NMR (270 MHz, DMSO-*d*₆, TSP) δ 1.13 (3H, d, *J*=7.3 Hz), 1.15 (3H, d, *J*=6.1 Hz), 2.10 (1H, dd, *J*=16.9 and 4.3 Hz), 2.77 (1H, dd, *J*=16.9 and 7.7 Hz), 3.08 (1H, dd, *J*=10.8 and 3.9 Hz), 3.20 (1H, dd, *J*=6.5 and 2.7 Hz), 3.15~3.60 (4H, m), 3.69 (1H, dd, *J*=10.8 and 7.5 Hz), 3.91~4.03 (2H, m), 4.17 (1H, dd, *J*=9.3 and 2.7 Hz), 5.03 (1H, br s), 7.82 (1H, s).

Preparation of Sodium (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*S*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**6n**)

i) 4-Nitrobenzyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-hydroxyethyl]-1-methyl-2-[(*S*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate **4n**

To a solution of 4-nitrobenzyl (1*R*,5*S*,6*S*)-2-diphenylphosphoryloxy-6-[(*R*)-1-hydroxyethyl]-1-methyl-1-carbapen-2-em-3-carboxylate **1** (939 mg, 1.58 mmol) was added a solution of the mercaptan **3n** (185 μ g, 1.58 mmol) in CH₃CN and diisopropylethylamine (330 μ l, 1.89 mmol). The mixture was stirred for 1.5 hours under ice-cooling and then left to stand overnight at 4°C. The mixture was diluted with EtOAc, washed with water and brine, and then concentrated under reduced pressure. The residue was chromatographed through silica gel (30 g) with EtOAc-MeOH (from 1:0 to 9:1) as the eluent to give the compound **4n** as a colorless powder

(654 mg, 1.42 mmol, 89% yield); IR (KBr) cm^{-1} 1771, 1701, 1522, 1347, 1208; ^1H NMR (270 MHz, DMSO- d_6 , TMS) δ 1.16 (3H, d, $J=6.0$ Hz), 1.18 (3H, d, $J=7.3$ Hz), 2.02 (1H, dd, $J=17.1$ and 4.9 Hz), 2.72 (1H, dd, $J=17.1$ and 8.3 Hz), 3.15 (1H, dd, $J=10.6$ and 3.3 Hz), 3.29 (1H, dd, $J=6.2$ and 2.5 Hz), 3.40~3.48 (1H, m), 3.74 (1H, dd, $J=10.7$ and 6.3 Hz), 3.94~4.05 (3H, m), 4.24 (1H, dd, $J=9.8$ and 2.9 Hz), 5.06 (1H, d, $J=4.9$ Hz), 5.30 (1H, d, $J=14.2$ Hz), 5.46 (1H, d, $J=14.2$ Hz), 7.71 (2H, d, $J=8.8$ Hz), 7.81 (1H, s), 8.23 (2H, d, $J=8.8$ Hz).

ii) Sodium (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*S*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate **6n**

To a solution of the compound **4n** (648 mg, 1.40 mmol) in a mixture of THF (33 ml) and 0.1 M aq. phosphate buffer (pH 7.0) (33 ml) was added 10% Pd-charcoal (700 mg), and the mixture was stirred vigorously at 22°C in an atmosphere of hydrogen for 1.5 hours. The catalyst was removed by filtration from the reaction mixture, and the filtrate was washed twice with Et₂O. The aqueous layer was then concentrated by evaporation under reduced pressure, and the resulting residue was chromatographed through Cosmosil 75C₁₈ PREP (60 ml) developed with water. The desired fraction was concentrated to 10 ml, and then lyophilized to give the title compound **6n** as a colorless powder (246 mg, 0.70 mmol, 50% yield);

Anal Calcd for C₁₄H₁₇N₂O₅SNa·2H₂O:

C 43.75, H 5.51, N 7.29, S 8.34.

Found: C 43.40, H 5.48, N 6.96, S 8.14.

IR (KBr) cm^{-1} 1725, 1683, 1591, 1387, 1372, 1302; UV (H₂O) nm 299; ^1H NMR (270 MHz, D₂O, TSP) δ 1.23 (3H, d, $J=7.2$ Hz), 1.30 (3H, d, $J=6.5$ Hz), 2.32 (1H, dd, $J=17.9$ and 4.4 Hz), 2.94 (1H, dd, $J=17.9$ and 8.6 Hz), 3.30~3.38 (1H, m), 3.40 (1H, dd, $J=11.1$ and 3.5 Hz), 3.46 (1H, dd, $J=6.2$ and 2.5 Hz), 3.88 (1H, dd, $J=11.1$ and 6.6 Hz), 4.05~4.10 (1H, m), 4.21~4.29 (2H, m).

Preparation of Sodium (5*R*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**7m**)

i) 4-Nitrobenzyl (5*R*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate **5m**

To a solution of 4-nitrobenzyl (5*R*,6*S*)-6-[(*R*)-1-hydroxyethyl]-2-oxo-1-carbapenam-3-carboxylate (2230 mg, 6.59 mmol) and diphenylphosphorylchloride (1.43 ml, 6.91 mmol) in CH₃CN (20 ml) was added diisopropyl-

ethylamine (1.31 ml, 7.54 mmol) under ice-cooling, and the mixture was stirred for 2 hours. To the mixture was added a solution of mercaptan **3m** (750 mg, 6.28 mmol) and diisopropylethylamine (1.31 ml, 7.54 mmol) in CH₃CN (20 ml). The mixture was stirred for 1 hour under ice-cooling. The crystalline compound which precipitated from the reaction mixture during the period was collected by filtration and dried to give the title compound **5m** as crystals (1.97 g, 4.40 mmol, 67% yield);

Anal Calcd for C₂₀H₂₁N₃O₇S:

C 53.69, H 4.73, N 9.39, S 7.17.

Found: C 53.87, H 4.69, N 9.32, S 7.09.

IR (KBr) cm^{-1} 1781, 1697, 1682, 1520, 1350, 1335, 1280, 1133; ^1H NMR (270 MHz, DMSO- d_6 , TMS) δ 1.14 (3H, d, $J=6.2$ Hz), 2.15 (1H, dd, $J=17.1$ and 5.2 Hz), 2.74 (1H, dd, $J=17.1$ and 8.4 Hz), 3.21 (1H, dd, $J=10.6$ and 4.1 Hz), 3.25~3.39 (2H, m), 3.69 (1H, dd, $J=10.6$ and 7.1 Hz), 3.91~4.21 (3H, m), 5.10 (1H, d, $J=5.0$ Hz), 5.30 (1H, d, $J=14.2$ Hz), 5.43 (1H, d, $J=14.2$ Hz), 7.70 (2H, d, $J=8.6$ Hz), 7.85 (1H, s), 8.23 (2H, d, $J=8.6$ Hz).

ii) Sodium (5*R*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate **7m**

To a solution of the compound **5m** (1000 mg, 2.23 mmol) and NaHCO₃ (187 mg, 2.23 mmol) in the mixture of THF (30 ml) and H₂O (25 ml) was added 10% Pd-charcoal (1000 mg), and the mixture was stirred vigorously at 30°C in an atmosphere of hydrogen for 1.5 hours. The catalyst was removed from the reaction mixture by filtration, and the filtrate was washed twice with EtOAc. The aqueous layer was then concentrated by evaporation under reduced pressure, and the resulting residue was chromatographed through Cosmosil 75C₁₈ PREP (20 g) developed with water. The desired fraction was concentrated to 10 ml, and then lyophilized to give the title compound **7m** as a colorless powder (622 mg, 1.86 mmol, 83% yield);

Anal Calcd for C₁₃H₁₅N₂O₅SNa·H₂O:

C 44.32, H 4.86, N 7.95.

Found: C 44.37, H 4.81, N 7.95.

IR (KBr) cm^{-1} 1755, 1686, 1593, 1394, 1296, 1252. ^1H NMR (400 MHz, D₂O, TSP) δ 1.29 (3H, d, $J=6.4$ Hz), 2.43 (1H, dd, $J=16.7$ and 4.9 Hz), 2.97 (1H, dd, $J=16.7$ and 8.6 Hz), 3.16 (1H, dd, $J=17.4$ and 8.6 Hz), 3.24 (1H, dd, $J=17.4$ and 9.6 Hz), 3.38~3.43 (2H, m), 3.89 (1H, dd, $J=11.3$ and 7.6 Hz), 4.09 (1H, m), 4.20~4.27 (2H, m).

Pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (11, CS-834)

To a solution of **6m** (4.76 g, 13.6 mmol) in *N,N*-dimethylacetamide (35 ml) was added pivaloyloxymethyl iodide (3.60 g, 14.8 mmol) under ice-cooling, and the mixture was stirred for 30 minutes. The mixture was diluted with EtOAc and washed with water and brine. The EtOAc layer was dehydrated using anhydrous Na₂SO₄, and then concentrated by evaporation under reduced pressure. The resulting residue in the form of an amorphous powder (4.54 g) was dissolved in EtOH-CH₂Cl₂ (1:1), and then the CH₂Cl₂ was evaporated under reduced pressure. Crystals were formed during the evaporation. The resulting crystals were collected by filtration and dried to give the title compound **11** (3.68 g, 8.35 mmol, 61% yield): mp 189°C;

Anal Calcd for C₂₀H₂₈N₂O₇S:

C 54.53, H 6.41, N 6.36, S 7.28.

Found: C 54.48, H 6.54, N 6.32, S 7.38.

IR (KBr) cm⁻¹ 3336, 1764, 1751, 1717, 1691, 1542, 1347, 1213, 1160, 1114, 995; UV (CH₃CN) nm 324; ¹H NMR (400 MHz, DMSO-*d*₆, TMS) δ 1.10~1.18 (15H, m), 2.11 (1H, dd, *J*=17.0 and 4.3 Hz), 2.78 (1H, dd, *J*=17.0 and 7.7 Hz), 3.09 (1H, dd, *J*=10.9 and 3.9 Hz), 3.25 (1H, dd, *J*=6.2 and 2.5 Hz), 3.44~3.48 (1H, m), 3.71 (1H, dd, *J*=10.9 and 7.6 Hz), 3.94~4.00 (1H, m), 4.04~4.09 (1H, m), 4.23 (1H, dd, *J*=9.5 and 2.5 Hz), 5.08 (1H, d, *J*=5.1 Hz), 5.73 (1H, d, *J*=5.9 Hz), 5.88 (1H, d, 5.9 Hz), 7.84 (1H, s).

1-Methylcyclohexylcarbonyloxymethyl (1*R*,5*S*,6*S*)-2-[(*R*)-5-Oxopyrrolidin-3-ylthio]-6-[(*R*)-1-hydroxyethyl]-1-methyl-1-carbapen-2-em-3-carboxylate (12)

Compound **12** was prepared from compound **6m** and 1-methylcyclohexylcarbonyloxymethyl iodide as described above. mp 160°C;

Anal Calcd for C₂₃H₃₂N₂O₇S:

C 57.48, H 6.71, N 5.83, S 6.67.

Found: C 57.25, H 6.86, N 5.80, S 6.38.

IR (KBr) cm⁻¹ 1761, 1687, 1341; UV (CH₃CN) nm 323; ¹H NMR (270 MHz, DMSO-*d*₆, TMS) δ 1.10 (3H, s), 1.13~1.30 (11H, m), 1.44~1.49 (3H, m), 1.89~1.93 (2H, m), 2.10 (1H, dd, *J*=16.8 and 4.4 Hz), 2.79 (1H, dd, *J*=16.8 and 7.8 Hz), 3.08 (1H, dd, *J*=11.0 and 3.7 Hz), 3.24 (1H, dd, *J*=16.8 and 2.5 Hz), 3.43~3.47 (1H, m), 3.68~3.73 (1H, m), 3.94~3.98 (1H, m), 4.05~4.08 (1H, m), 4.21 (1H, dd, *J*=9.5 and 2.2 Hz), 5.08 (1H, brs), 5.76 (1H, d, *J*=5.7 Hz), 5.87 (1H, d, *J*=5.7 Hz), 7.84 (1H, s).

Cyclohexyloxycarbonyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (13)

Compound **13** was prepared from compound **6m** and cyclohexyloxycarbonyloxymethyl iodide as described above.

IR (KBr) cm⁻¹ 1763, 1695, 1277; ¹H NMR (270 MHz, DMSO-*d*₆, TMS) δ 1.13~1.17 (6H, m), 1.18~1.52 (6H, m), 1.58~1.70 (2H, m), 1.80~1.90 (2H, m), 2.14 (1H, dd, *J*=17.0 and 4.4 Hz), 2.78 (1H, dd, *J*=17.0 and 7.8 Hz), 3.10 (1H, dd, *J*=10.7 and 3.9 Hz), 3.25 (1H, dd, *J*=6.3 and 2.9 Hz), 3.42~3.55 (1H, m), 3.71 (1H, dd, *J*=11.0 and 7.8 Hz), 3.90~4.00 (1H, m), 4.02~4.13 (1H, m), 4.23 (1H, dd, *J*=9.3 and 2.4 Hz), 4.53~4.62 (1H, m), 5.07 (1H, d, *J*=5.4 Hz), 5.72 (1H, d, *J*=6.3 Hz), 5.83 (1H, d, *J*=6.3 Hz), 7.84 (1H, s).

1-(Isopropoxycarbonyloxy) ethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (14)

Compound **14** was prepared from compound **6m** and 1-(isopropoxycarbonyloxy) ethyl iodide as described above.

IR (KBr) cm⁻¹ 1759, 1693, 1273, 1071; ¹H NMR (270 MHz, DMSO-*d*₆, TMS) δ 1.12~1.17 (6H, m), 1.20~1.26 (6H, m), 1.44~1.48 (3H, m), 2.08~2.20 (1H, m), 2.79 (1H, dd, *J*=17.1 and 7.8 Hz), 3.10 (1H, dd, *J*=11.2 and 3.9 Hz), 3.21~3.26 (1H, m), 3.39~3.50 (1H, m), 3.71 (1H, dd, *J*=10.7 and 7.8 Hz), 3.90~4.00 (1H, m), 4.01~4.10 (1H, m), 4.19~4.20 (1H, m), 4.70~4.85 (1H, m), 5.05~5.08 (1H, m), 6.66~6.75 (1H, m), 7.83 (1H, s).

Pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-1-methyl-2-[(*S*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (15)

Compound **15** was prepared from compound **6n** and pivaloyloxymethyl iodide as described above. IR (KBr) cm⁻¹ 1756, 1700, 1346, 1280, 1116; ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.23 (9H, s), 1.29 (3H, d, *J*=7.1 Hz), 1.34 (3H, d, *J*=6.4 Hz), 2.33 (1H, dd, *J*=17.5 and 6.0 Hz), 2.78 (1H, dd, *J*=17.5 and 8.7 Hz), 3.25~3.33 (2H, m), 3.38 (1H, dd, *J*=10.0 and 4.5 Hz), 3.80 (1H, dd, *J*=10.0 and 7.2 Hz), 3.96~4.05 (1H, m), 4.22~4.28 (2H, m), 5.78 (1H, brs), 5.83 (1H, d, *J*=5.7 Hz), 5.97 (1H, d, 5.7 Hz).

Pivaloyloxymethyl (5*R*,6*S*)-6-[(*R*)-1-Hydroxyethyl]-2-[(*R*)-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (16)

Compound **16** was prepared from compound **7m** and pivaloyloxymethyl iodide as described above. mp 189~190°C; IR (KBr) cm^{-1} 1772, 1759, 1701, 1540, 1336, 1280, 1202, 1157, 1111; ^1H NMR (270 MHz, DMSO- d_6 , TMS) δ 1.14 (9H, s), 2.14 (1H, dd, $J=17.0$ and 5.2 Hz), 2.74 (1H, dd, $J=17.0$ and 8.2 Hz), 3.22 (1H, dd, $J=10.6$ and 4.1 Hz), 3.69 (1H, dd, $J=10.6$ and 7.3 Hz), 3.89~4.00 (1H, m), 4.02~4.20 (2H, m), 5.09 (1H, d, $J=4.8$ Hz), 5.74 (1H, d, $J=5.8$ Hz), 5.84 (1H, d, $J=5.8$ Hz), 7.85 (1H, s).

Measurement of Antibacterial Activity

MICs were measured on Nutrient agar (Eiken Chemical Ltd.) by the twofold dilution method. The inoculum size of the bacteria was one-loopful of 10^7 cfu/ml.

Stability of Carbapenems against Renal DHP-I of Mice

The relative hydrolysis rate was determined by using 20% homogenate of kidney cortex from mice. An equi-volume of homogenate was added to 50 mM MOPS buffer (pH 7.0) which containing compounds of tested (2 mM, final concentration: 1 mM) and the mixture was incubated at 37°C. At appropriate time intervals, portion of mixture was transferred to another test tube that contained 3-times volume of methanol to stop enzymatic reaction. After vortex and centrifugation, the supernatants were injected to HPLC. The HPLC conditions are as follows: Column; A-312 ODS (6 mm i.d. \times 150 mm length) from YMC, eluent; 20 mM MES buffer (pH 6.0)-methanol (8:2), flow rate; 1.0 ml/minute and UV detection at 300 nm. The hydrolysis rates of compounds were determined from plots of peak areas of the compounds against time of samplings and expressed as relative hydrolysis rate against compound **6m**. DHP-I activity of the homogenate was confirmed by the method of CAMPBELL²³) using glycyldehydrophenylalanine as a substrate.

Urinary Recovery of Carbapenems in Mice

i) Administration

Carbapenems **6a** ~ **n** and **7m** (dose: 50 mg/kg as sodium salt) were dissolved in water and then subcutaneously administered to mice (n=5, SPF *ddY* strain). A mixture of cilastatin sodium and carbapenems **6m**, **6n** and **7m**

(dose: 50 mg/kg each as sodium salt) was dissolved in water and then subcutaneously administered to mice (n=5, SPF *ddY* strain). Carbapenem esters **11** ~ **16** (dose: 50 mg/kg as **6m**, **6n** and **7m**) suspended in 0.5% tragacanth were orally administered to mice (n=5, SPF *ddY* strain).

ii) Determination of Urinary Recovery

Urine was collected at 8 hours and 24 hours after administration. Excretion as the parent carbapenem was determined by bioassay using *Bacillus subtilis* ATCC 6633. Urinary recovery (% , 0~24 hours) was calculated based on the excretion and the initial dose.

Pharmacokinetic Parameters of Carbapenems in Dogs

i) Administration

Carbapenem **6m** (dose: 10 mg/kg) dissolved in water was intravenously administered to dogs (n=3, Beagle). Carbapenem esters **11** and **12** (dose: 10 mg/kg as **6m**) dissolved in dimethylacetamide - polyethyleneglycol 400 - water (1:4:5) were orally administered to dogs (n=3, Beagle).

ii) Determination of Pharmacokinetic Parameters

The concentration of R-95867 in blood was measured by HPLC method at suitable intervals after administration. Pharmacokinetic parameters were determined by the simulation program.

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